

Hymenoptera functional groups' shifts in disturbance gradients at Andean forests in Southern Ecuador

Marina Mazón^{1,2}, Ximena López³, Oscar Romero⁴

I Biodiversity and Ecosystem Services Research Program, Universidad Nacional de Loja, Ciudadela Universitaria, sector La Argelia, EC 110101 Loja, Ecuador 2 Departamento de Ciencias Ambientales y Recursos Naturales, Universidad de Alicante, Apdo. Correos 99, 03080 Alicante, Spain 3 Carrera de Ingeniería en Manejo y Conservación del Medio Ambiente, Universidad Nacional de Loja, Ciudadela Universitaria, sector La Argelia, EC 110101 Loja, Ecuador

Corresponding author: Marina Mazón (marinamazonmor@gmail.com)

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Abstract

Ecosystems under ecological restoration should be monitored in order to investigate if the ecosystem is being functionally recovered, especially in highly vulnerable biodiversity hotspots like Andean forests. Here we sampled Hymenoptera families in four Andean forest reserves above 1800 masl from Southern Ecuador, in three conservation levels in each forest: low (degraded), medium (10–15 years of recovery) and high (well-conserved forest). All Hymenoptera families were classified into four functional groups: predators, herbivores, pollinators and parasitoids. A total of 32 hymenopteran families were collected, with parasitoids clearly dominating in the samples. Family assemblages were not statistically different, neither in abundance nor family richness. Assemblages were more similar between them in the high and medium areas than in low conservation areas, where assemblages were very variable and showed a higher functional diversity in two of the reserves. The low presence of pollinators may be due to the high humidity during the sampling and the sampling method. Although some results are promising for the restoring trajectory, especially for parasitoids, we should keep in mind that this is at family level, so it would be interesting to know if these patterns persist at lower taxonomic levels.

Keywords

Herbivores, Hymenoptera families, Parasitoids, pollinators, predators

Introduction

Ecosystems provide a wide range of services that are at risk because of biodiversity loss derived from demographic growth and land use changes (De Groot et al. 2012). Ecosystems do have an ability to recover previous biodiversity up to a certain point, beyond which they need some assistance. The process of "assisting the recovery of an ecosystem that has been degraded, damaged or destroyed" is called ecological restoration (SER 2004). However, since ecosystems may progress in different ways after restoration efforts, depending on many different environmental variables (Bullock et al. 2011), monitoring is essential to know how ecosystem attributes are recovering.

Andean forests are considered as a highly vulnerable biodiversity hotspot (Myers et al. 2000; Mathez-Stiefel et al. 2017; Murcia et al. 2017), with most of its plant species already experiencing shifts in their distribution because of global warming (Fadrique et al. 2018). Although there are some initiatives for restoring Andean forests, they are not being monitored comprehensively (Mazón et al. 2019), with plant structure and diversity being evaluated in most cases (Yepes et al. 2010; Szejner et al. 2011; Camelo et al. 2017). Therefore, little is known about whether many other ecosystem services are being recovered in these forests.

Hymenoptera is one of the most important and diverse insect orders, including representative groups of two of the most essential ecosystem services: pollination (i.e., bees) and natural pest control (i.e., parasitoid wasps). Both pollinators and parasitoids (whose hosts are mostly herbivorous insects), because of their close relationship with plants, have been used as indicators for ecosystem disturbances (Vos et al. 2001; Requier 2019) and their recovery (Maeto et al. 2008; Ferronato et al. 2018). Pollinators have received much attention in recent years because of their worrying decreasing populations and their high vulnerability to climate change (Potts et al. 2010; Powney et al. 2019). However, other hymenopterans like parasitoid wasps and ants are also sensitive to ecosystem degradation and fragmentation (de Sassi and Tylianakis 2012). Since hymenopteran families may have different responses to habitat degradation (Banks et al. 2013), they may also have different responses when being monitored in ecological restoration projects, as well as the roles they play in the ecosystem. For instance, bee and parasitoid diversity has been shown to be higher in restored areas than in disturbed ones (Barbieri Junior and Penteado-Dias 2012; Marrec et al. 2018) but the species interaction and functions appear to take a longer time to recover (Albrecht et al. 2007). Therefore, species diversity per se may not be the best predictor of ecosystem services, but, rather, diversity at the functional level (Griffin et al. 2013). In consequence, measuring functional diversity may predict ecosystem functioning better than the traditional species diversity measures (Loreau et al. 2001).

In this study we aim to 1) identify the Hymenoptera families present in different disturbance levels in Andean forests, and 2) evaluate how functional groups within Hymenoptera are shifting as disturbance increases.

Methods

Study area

The research was done in four protected areas of Andean forest located in Loja and Zamora Chinchipe provinces, in the buffer area of Podocarpus National Park, at southern Ecuador (Fig. 1), with altitudes ranging 1840–2630 masl (Table 1). Andean forests are ecosystems located between 1200 and 3600 masl at Los Andes mountains (Quintero et al. 2017), characterized by their elevated humidity and a high diversity of ferns, lichens, mosses, palms and orchids (Bravo 2014; Kattan 2017).

Sampling and identification

In each reserve, we identified three areas of different conservation status with the help of their personal staff: a well–conserved area, an area under about 10–15 years of recovery, either naturally or assisted, and a degraded area (Table 1). In each area, we installed a white Townes style Malaise trap (Townes 1972) for six weeks, from December 2015 to January 2016 in most reserves, except for Tapichalaca, which was sampled from March to April 2018; in all cases, sampling was carried out during the rainy season. We filled pots with ethanol 70% and replaced them every two weeks, i.e., having three samples per area.

We sorted and identified all hymenopteran specimens to family level, and then we classified them into four major functional groups: predators, herbivores, pollinators and parasitoids, following Fernández and Sharkey (2006).

Data analyses

We compared the family assemblages in the three conservation levels by a non-metric multidimensional scaling (NMDS) and a PERMANOVA with 9999 permutations, using Jaccard index for similarity, which considers presence/absence of families, regardless of the relative abundances of each one. We did the same analyses to check for differences in the hymenopteran families assemblages related to the reserves.

Regarding the functional groups, we calculated both richness (i.e., number of families) and abundance (i.e., number of individuals) for every functional group, and we compared them across the conservation levels by means of a KRUSKAL-WALLIS test and a post-hoc DUNN test.

Additionally, we evaluated functional diversity with the Shannon index, considering the abundance of individuals belonging to every functional group. Since some samples were damaged and lost, we considered the mean abundance values for the two or three samples in every sampling site. Then, we compared Shannon indices in two ways: by the KRUSKAL-WALLIS with the four reserves as replicates, and in the four reserves treated as independent samples, compared by a randomization test with 1000 random partitions (Solow 1993).

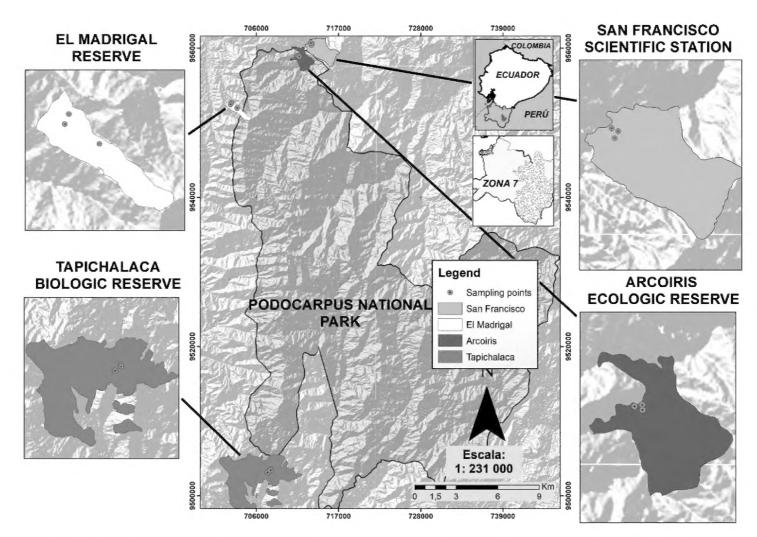


Figure 1. Localities of the four Andean forest reserves and the sampling points where the Malaise traps were placed in each one for collecting Hymenoptera.

The NMDS, PERMANOVA, KRUSKAL-WALLIS and post-hoc tests were run with software Past version 3.0 (Hammer et al. 2001). Shannon diversity index and randomization tests were performed with software Species Diversity and Richness 3.02 (Pisces Conservation, Ltd., Pennington, Lymington, UK).

Results

We collected a total of 32 hymenopteran families, mostly belonging to the parasitoid functional group, with family Ichneumonidae clearly dominating the sampling (1930 individuals). We only collected seven specimens belonging to three families from the pollinator functional group (Table 2).

Families assemblages were not statistically different, neither regarding the conservation level (F = 1.354, p = 0.156) nor the reserve (F = 1.382, p = 0.118). In the NMDS, although the samples were distributed from high to low conservation level (Fig. 2A), those from the low level were the most variable, especially the sample from Madrigal reserve, which was more different than the others in terms of hymenopteran family composition (Fig. 2B). In both cases, axis 1 explained more of the data (50.14% and 49.87%, respectively) than axis 2.

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Reserve	Conservation level	Coordinates X	Coordinates Y	Altitud (masl)
ECSF	high	-79.077	-3.973	ca 1870
	medium	-79.078	-3.975	ca 1860
	low	-79.079	-3.972	ca 1840
Arcoíris	high	-79.095	-3.988	ca 2160
	medium	-79.093	-3.988	ca 2160
	low	-79.093	-3.989	ca 2160
Madrigal	high	-79.168	-4.051	ca 2520
	medium	-79.175	-4.045	ca 2350
	low	-79.176	-4.047	ca 2400
Tapichalaca	high	-79.126	-4.489	ca 2570
	medium	-79.130	-4.493	ca 2520
	low	-79.126	-4.490	ca 2620

Table 2. Abundances and functional groups of every collected hymenopteran family in the three conservation levels (high, medium and low) from the four Andean forest reserves in southern Ecuador. PRED = predators, HERB = herbivores, PAR = parasitoids, POL = pollinators.

Family	Functional groups		ECSF			Arcoiris			Madrigal		Г	apichalaca	ı	Total
		high	medium	low	high	medium	low	high	medium	low	high	medium	low	
Pompilidae	PRED	0	0	0	1	0	0	0	0	0	1	5	1	8
Sphecidae	PRED	0	0	0	0	0	0	0	0	0	0	1	0	1
Vespidae	PRED	1	0	0	0	1	4	1	1	0	0	14	5	27
Formicidae	PRED	19	4	4	24	2	1	3	1	0	0	2	3	65
Pergidae	HERB	0	0	0	0	0	0	4	0	0	1	1	0	6
Tenthredinidae	HERB	0	0	0	2	0	0	3	0	0	1	2	4	12
Xiphydriidae	HERB	0	0	0	2	1	0	0	0	0	2	0	0	5
Bethylidae	PAR	10	0	0	1	0	1	1	1	0	0	1	6	21
Braconidae	PAR	167	9	12	60	15	6	134	25	2	38	31	76	577
Diapriidae	PAR	48	1	1	6	1	1	14	3	0	6	17	36	134
Dryinidae	PAR	0	1	1	0	0	0	0	1	0	2	10	4	19
Embolemidae	PAR	2	0	0	0	0	0	0	0	0	0	0	0	2
Eucharitidae	PAR	0	0	0	0	1	0	0	0	0	0	0	0	1
Eulophidae	PAR	5	0	0	0	0	0	1	0	0	1	2	2	11
Eupelmidae	PAR	0	1	1	0	0	0	0	0	0	1	1	0	4
Eurytomidae	PAR	3	0	0	0	0	0	0	0	0	0	0	0	3
Evaniidae	PAR	25	2	0	14	4	2	11	3	0	1	14	9	92
Figitidae	PAR	1	0	0	2	0	0	3	0	0	4	2	5	17
Ichneumonidae	PAR	300	33	6	188	72	23	248	101	6	195	233	401	1930
Liopteridae	PAR	0	0	0	0	0	0	0	0	0	0	2	0	2
Mutillidae	PAR	5	0	0	0	0	0	0	0	0	0	0	0	5
Mymaridae	PAR	0	0	0	0	0	0	0	0	0	0	0	1	1
Orussidae	PAR	0	0	0	0	1	0	0	1	0	2	1	0	5
Perilampidae	PAR	3	0	0	0	0	0	1	0	0	0	0	0	4
Platygastridae	PAR	13	2	2	5	0	0	3	0	0	5	6	8	44
Proctotrupidae	PAR	2	0	0	0	3	0	0	0	0	2	1	2	10
Pteromalidae	PAR	6	0	0	2	0	0	1	0	0	0	6	0	16
Sapygidae	PAR	1	0	0	1	4	1	0	1	0	0	1	0	9
Tiphiidae	PAR	5	0	0	0	0	0	0	3	0	0	12	3	23
Agaonidae	POL	0	0	0	1	0	0	0	0	0	0	0	0	1
Apidae	POL	0	1	1	0	0	1	0	0	0	0	0	1	4
Halictidae	POL	0	0	0	1	0	0	0	0	0	0	2	0	3

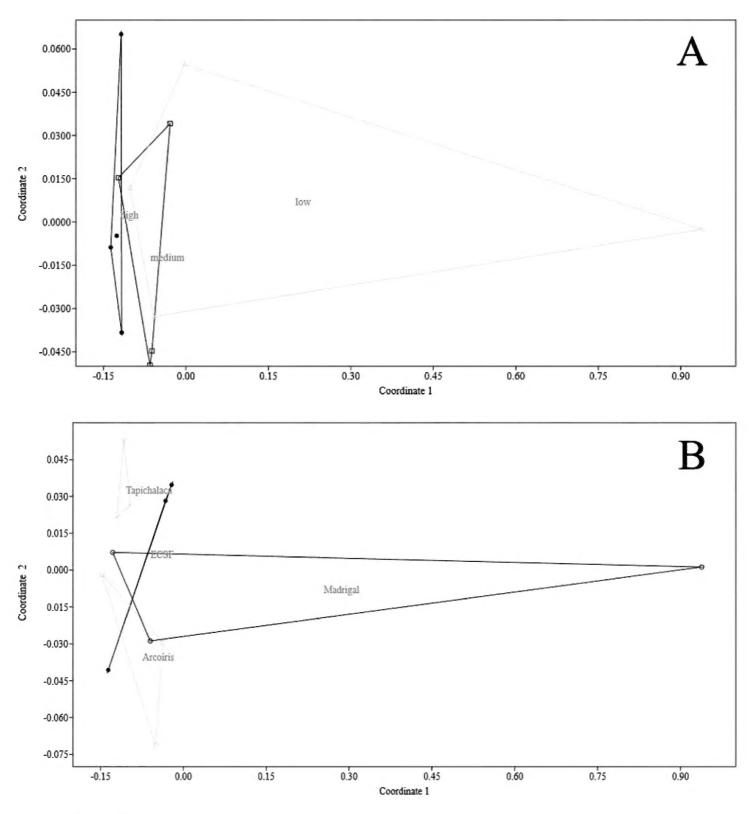


Figure 2. NMDS plots for hymenopteran families assemblages grouped by conservation level (**A**) and by reserve (**B**). Samples from the same group are gathered by convex hulls.

When comparing mean abundance and total richness of every functional group with the conservation levels no significant differences were found either (Table 3).

Regarding functional diversity, no significant results were obtained in the KRUSKAL-WALLIS test (H = 0.3462, p = 0.841). However, when treating every reserve independently, permutational tests gave very different results in the four reserves (Fig. 3). Surprisingly, functional diversity was significantly higher in the more perturbed areas in ECSF and Arcoiris. However, diversity in the areas under restoration (i.e., medium conservation level) was more similar to those in the conserved areas than in the highly perturbed ones.

Table 3. Results from Kruskal-Wallis test (H) and p-value (p) when comparing number of families (S) and number of individuals (Ab) from all Hymenoptera and every functional group (PRED = predators, HERB = herbivores, PAR = parasitoids, POL = pollinators) in the three conservation levels of Andean forests.

	Н	p
S_total Hymenoptera	3.298	0.19
Ab_total Hymenoptera	3.962	0.138
S_PRED	0.183	0.903
Ab_PRED	0.269	0.872
S_HERB	3.010	0.174
Ab_HERB	3.151	0.207
S_PAR	3.537	0.171
Ab_PAR	3.962	0.138
S_POL	0.644	0.671
Ab_POL	0.5	0.74

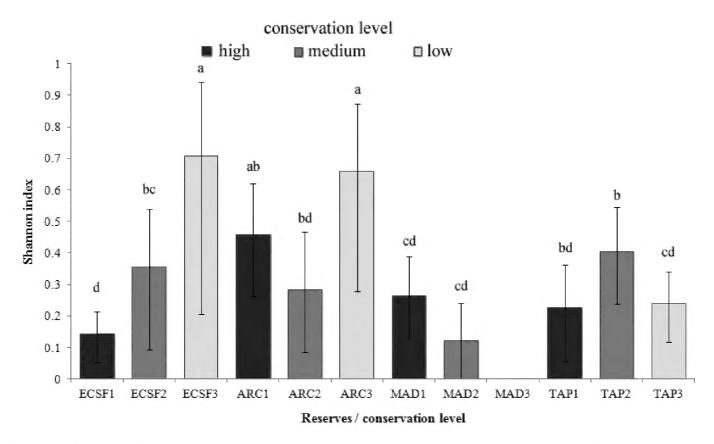


Figure 3. Mean values of Shannon index for functional diversity of Hymenoptera collected in the three conservation levels (high, medium, low) from the four reserves of Andean forests. Vertical bars denote 95% confidence intervals. Different letters indicate statistically significant differences.

Discussion

In the present research we found 32 hymenopteran families, which represents 36.31% of Neotropical families (Fernández and Sharkey 2006). Ichneumonidae and Braconidae were the most abundant, which are usually amongst the main hymenopteran families collected by Malaise trap in many ecosystems, as in preserved sandbanks (Oliveira et al. 2009), open pastures (Castiglioni et al. 2017) or cacao plantations (Mazón 2016). The high abundance of these two families is also reflected in the dominance registered for parasitoid functional groups. Actually, parasitoids are the most abundant and diverse hymenopteran group (Fernández and Sharkey 2006) and are able to adapt

to different environmental conditions, with a rather wide range of potential hosts (Santos and Quicke 2011).

However, the low occurrence of hymenopteran pollinators was unexpected. Bees are abundant in Andes even above 2500 masl (Gonzalez and Engel 2004), and at least 51 bee species have been recorded from Southern Ecuador (Rasmussen 2004). Since one of the main factors limiting bee nesting is high humidity (Michener 2007; Reyes-Novelo et al. 2009), the season when sampling was done may have influenced these results. Furthermore, Malaise traps may not be the most appropriate sampling method for bees. Although showed as highly effective for parasitoids (Mazón and Bordera 2008) and aculeates in general (Volpato et al. 2020), with some reported bias either towards males (Aguiar and Santos 2010) or females (Mazón et al. 2020), scent-baited traps or entomological nets seem to work better for bees (Santos Júnior et al. 2014; Ferronato et al. 2018; Alvarenga et al. 2020). Other sampling methods should be used to complement Malaise traps, especially when time for field work is brief (McGravy et al. 2016; Saunders and Ward 2018). Sampling should also include different weather seasons in order to have a better representation of all Hymenoptera families and to clarify if bees have reduced populations in these forests.

No differences among conservation levels were found. Restoration may favour the presence of wild bees (Araújo et al. 2018; Taki et al. 2018; Alvarenga et al. 2020) and parasitoids (Marrec et al. 2018), but some hymenopteran assemblages from restored areas may be functionally similar to those from forests (Montoya-Pfeiffer et al. 2020), and their diversity has been found to be lower in forests compared to more open areas (Pardo and Gonzalez 2007). Therefore, it seems that, even when the areas labeled as low conservation presented a high degree of perturbation, being embedded into a preserved area matrix helped to increase diversity in these areas, since the hymenopteran diversity is positively affected by the proximity to forest (Banks et al. 2013). Furthermore, not all types of forest restoration will equally favour Hymenopterans, with ecological restoration, as applied in all the studied reserves, being more effective than monoculture tree plantations (de Araújo et al. 2019). This indicates that ecological restoration in these areas has a positive effect on hymenopteran diversity, and the more disturbed areas within the reserve matrix do not significantly impact these assemblages, at least at family level.

Another aspect that may have masked the effect of conservation level on Hymenoptera richness and abundance are the altitudinal differences amongst some of the reserves. Parasitoids (van Noort 2004; Veijalainen et al. 2014; Hall et al. 2015), wasps and bees (Perillo et al. 2017; Widhiono et al. 2017) and ants (Guerrero and Sarmiento 2010; Burwell and Nakamura 2011) have been found to be sensitive to altitude in tropical and subtropical ecosystems. However, to see more clearly how Hymenoptera are responding to both altitude and conservation level it would be necessary to use lower taxa. Higher taxa have been used as surrogates for species to assess biodiversity in highly species-rich ecosystems or when sampling time is limited. For this purpose, genus (Derraik et al. 2010; Vieira et al. 2012) or subfamily level (Mazón 2016) have proved to be useful surrogates in Hymenoptera. Although family level may not be such

a good surrogate for species diversity than other lower taxonomic levels (Balmford et al. 1996), it can provide important ecological information, especially when grouped into functional groups (Bragança et al. 1998).

When looking at assemblage composition, samples were grouped according to conservation level rather than to reserves, although samples were very separated one from another, showing that assemblages, even in the same conservation level, were highly variable. The type of disturbance may be influencing these results, since it will affect the way the ecosystem responds to it (Jones and Schmitz 2009; Pandit et al. 2018), and therefore insect communities will also change accordingly. However, that variable was not considered in this study, so it is difficult to draw those conclusions. On the other hand, most of the assemblages from conserved areas were very similar amongst themselves except for the Tapichalaca reserve, that was clearly separate from the others, indicating the Andes may be acting as an ecological barrier for these insects, even at a local scale.

Regarding overall functional diversity, it was significantly higher in the low conservation areas of two reserves. Although it can not be seen as a consistent pattern, open areas may serve as corridors for insects (Mazón and Bordera 2014; Gutiérrez-Chacón et al. 2020) connecting well-conserved forests, and therefore these more disturbed areas would have more diverse assemblages. Also, by being embedded in a conserved matrix, these areas may be acting as ecotones, which may harbor a higher diversity of Hymenoptera than the forests, as seen for ants (Coelho and Ribeiro 2006), bees (Polatto and Alves 2020; Theodorou et al. 2020) and parasitoids (Rossetti et al. 2013). However, the occurrence of insects in these ecotones would be limited by some traits as the presence of trees (especially for nesting bees and wasps, Polatto and Alves 2020) or the dispersal ability of each species (Hamm and Drossel 2017), therefore functional diversity based on the genus or species level, with more detailed functional groups, would provide a better information.

Ecological restoration is an effective way to recover the structure and function of ecosystems, but comprehensive monitoring should be carried out in order to investigate if its functions are becoming similar to those from a healthy well conserved reference ecosystem. Here we show promising results for parasitoid wasps, but not so for bees. It would be interesting to see what happens when identification goes to genus or species level, and whether trends observed for families are persisting for these lower taxonomic levels.

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References

- Aguiar AP, Santos BF (2010) Discovery of potent, unsuspected sampling disparities for malaise and Möricke traps, as shown for Neotropical Cryptini (Hymenoptera, Ichneumonidae). Journal of Insect Conservation 14: 199–206. https://doi.org/10.1007/s10841-009-9246-x
- Albrecht M, Duelli P, Schmid B, Müller CB (2007) Interaction diversity within quantified insect food webs in restored and adjacent intensively managed meadows. Journal of Animal Ecology 76: 1015–1025. https://doi.org/10.1111/j.1365-2656.2007.01264.x
- Alvarenga AS, Silveira FA, Júnior JES, Novais SMA, Quesada M, Neves FS (2020) Vegetation composition and structure determine wild bee communities in a tropical dry forest. Journal of Insect Conservation 24: 487–498. https://doi.org/10.1007/s10841-020-00231-5
- Araújo GJ, Monteiro GF, Messias MCTB, Antonini Y (2018) Restore it, and they will come: trap-nesting bee and wasp communities (Hymenoptera: Aculeata) are recovered by restoration of riparian forests. Journal of Insect Conservation 22(2): 245–256.
- Balmford A, Green MJB, Murray MG (1996) Using higher-taxon richness as a surrogate for species richness: I. Regional tests. Proceedings of the Royal Society London 263: 1267–1274. https://doi.org/10.1098/rspb.1996.0186
- Banks JE, Hannon L, Hanson P, Dietsch T, Castro S, Urena N, Chandler M (2013) Effects of proximity to forest habitat on hymenoptera diversity in a Costa Rican coffee agroecosystem. The Pan-Pacific Entomologist 89(1): 60–68. https://doi.org/10.3956/2012-28.1
- Barbieri Junior CA, Penteado-Dias AM (2012) Braconidae (Hymenoptera) fauna in native, degraded and restoration areas of the Vale do Paraíba, Sao Paulo state, Brazil. Brazilian Journal of Biology 72: 305–310. https://doi.org/10.1590/S1519-69842012000200011
- Bragança MAL, Zanuncio JC, Picanço M, Laranjeiro AJ (1998) Effects of environmental heterogeneity on Lepidoptera and Hymenoptera populations in Eucalyptus plantations in Brazil. Forest Ecology and Management 103: 287–292. https://doi.org/10.1016/S0378-1127(97)00226-0
- Bravo E (2014) La Biodiversidad en el Ecuador. Universidad Politécnica Salesiana. Cuenca, Ecuador.
- Bullock JM, Aronson J, Newton AC, Pywell RF, Rey-Benayas JM (2011) Restoration of ecosystem services and biodiversity: conflicts and opportunities. Trends in Ecology & Evolution 26(10): 541–549. https://doi.org/10.1016/j.tree.2011.06.011
- Burwell CJ, Nakamura A (2011) Distribution of ant species along an altitudinal transect in continuous rainforest in subtropical Queensland, Australia. Memoirs of the Queensland Museum 55(2): 391–411.

- Camelo OJ, Urrego LE, Orrego SA (2017) Environmental and socioeconomic drivers of woody vegetation recovery in human-modified landscape in the Rio Grande Basin (Colombian Andes). Restoration Ecology 25(6): 912–921. https://doi.org/10.1111/rec.12514
- Castiglioni E, Perioto NW, Lara IRL, Burla JP, Arbulo N, Aldabe J (2017) Análisis de esfuerzo de muestreo de himenópteros parasitoides en tres ambientes del este uruguayo. Innotec 13: 98–105. https://doi.org/10.26461/13.10
- Coelho IR, Ribeiro SP (2006) Environment heterogeneity and seasonal effects in ground-dwelling ant (Hymenoptera: Formicidae) assemblages in the Parque Estadual do Rio Doce, MG, Brazil. Neotropical Entomology 35: 19–29. https://doi.org/10.1590/S1519-566X2006000100004
- De Araújo GJ, Storck-Tonon D, Izzo TJ (2019) Is planting trees enough? The effect of different types of reforestation on the offspring of *Trypoxylon* (*Trypargillum*) lactitarse (Hymenoptera: Crabronidae) in the Southern Amazon. Neotropical Entomology 48: 572–582. https://doi.org/10.1007/s13744-019-00682-9
- De Groot R, Brander L, van der Ploeg S, Costanza R, Bernard F, Braat L, Christie M, Crossman N, Ghermandi A, Hein L, Hussain H, Kumar P, McVittie A, Portela R, Rodriguez LC, ten Brink P, van Beukering P (2012) Global estimates of the value of ecosystems and their services in monetary units. Ecosystem Services 1(1): 50–61. https://doi.org/10.1016/j.ecoser.2012.07.005
- De Sassi C, Tylianakis JM (2012) Climate change disproportionately increases herbivore over plant or parasitoid biomass. PLoS ONE 7(7): e40557. https://doi.org/10.1371/journal.pone.0040557
- Derraik JGB, Closs GP, Dickinson KJM (2010) Morphospecies and taxonomic species comparison for Hymenoptera. Journal of Insect Science 10: 108. https://doi.org/10.1673/031.010.10801
- Fadrique B, Báez S, Duque A, Malizia A, Blundo C, Carilla J, Osinaga-Acosta O, Malizia L, Silman M, Farfán-Ríos W, Malhi Y, Young KR, Cuesta F, Homeier J, Peralvo M, Pinto E, Jadan O, Aguirre N, Aguirre Z, Feeley KJ (2018) Widespread but heterogeneous responses of Andean forests to climate change. Nature 564: 207–212. https://doi.org/10.1038/s41586-018-0715-9
- Fernández F, Sharkey MJ (2006) Introducción a los Hymenoptera de la región neotropical. Sociedad Colombiana de Entomología y Universidad Nacional de Colombia, Bogotá D.C., 894 pp.
- Ferronato MCF, Giangarelli DC, Mazzaro D, Uemura N, Sofia SH (2018) Orchid bee (Apidae: Euglossini) communities in Atlantic forest remnants and restored areas in Paraná state, Brazil. Neotropical Entomology 47: 352–361. https://doi.org/10.1007/s13744-017-0530-2
- Gonzalez VH, Engel MS (2004) The Tropical Andean bee fauna (Insecta: Hymenoptera: Apoidea), with examples from Colombia. Entomologische Abhandlungen 62(1): 65–75.
- Griffin J, Byrnes J, Cardinale B (2013) Effects of predator richness on prey suppression: a meta-analysis. Ecology 94: 2180–2187. https://doi.org/10.1890/13-0179.1
- Guerrero RJ, Sarmiento CE (2010) Distribución altitudinal de hormigas (Hymenoptera, Formicidae) en la vertiente noroccidental de la Sierra Nevada de Santa Marta (Colombia). Acta Zoológica Mexicana 26(2): 279–302. https://doi.org/10.21829/azm.2010.262699

- Gutiérrez-Chacón C, Valderrama-A C, Klein A-M (2020) Biological corridors as important habitat structures for maintaning bees in a tropical fragmented landscape. Journal of Insect Conservation 24(1): 187–197. https://doi.org/10.1007/s10841-019-00205-2
- Hall CR, Burwell CJ, Nakamura A, Kitching RL (2015) Altitudinal variation of parasitic Hymenoptera assemblages in Australian subtropical rainforest. Austral Entomology 54(3): 246–258. https://doi.org/10.1111/aen.12114
- Hamm M, Drossel B (2017) Habitat heterogeneity hypothesis and edge effects in model metacommunities. Journal of Theoretical Biology 426: 40–48. https://doi.org/10.1016/j.jtbi.2017.05.022
- Hammer O, Harper DAT, Ryan PD (2001) Past: Paleontological Statistics software package for education and data analysis. Palaeontologia Electronica 4(1): 1–9.
- Jones HP, Schmitz OJ (2009) Rapid recovery of damaged ecosystems. PLoS ONE 4(5): e5653. https://doi.org/10.1371/journal.pone.0005653
- Kattan G (2017) Bosques Andinos y Subandinos del Departamento del Valle del Cauca, Colombia. Programa Colombia de Wildlife Conservation Society. Santiago de Cali, Valle del Cauca, Colombia.
- Loreau M, Naeem S, Inchausti P, Bengtsson J, Grime JP, Hector A, Hooper DU, Huston MA, Raffaelli D, Schmid B, Tilman D, Wardle DA (2001) Biodiversity and ecosystem functioning: current knowledge and future challenges. Science 294: 804–808. https://doi.org/10.1126/science.1064088
- Maeto K, Noerdjito WA, Belokobylskij SA, Fukuyama K (2008) Recovery of species diversity and composition of braconid parasitic wasps after reforestation of degraded grasslands in lowland East Kalimantan. Journal of Insect Conservation 13: 245–257. https://doi.org/10.1007/s10841-008-9164-3
- Marrec R, Pontbriand-Paré O, Legault S, James PMA (2018) Spatiotemporal variation in drivers of parasitoid metacommunity structure in continuous forest landscapes. Ecosphere 9(1): e02075. https://doi.org/10.1002/ecs2.2075
- Mathez-Stiefel S-L, Peralvo M, Báez S (2017) Hacia la conservación y la gobernanza sostenible de los paisajes de bosques andinos: Una agenda de investigación. Programa Bosques Andinos de la Agencia Suiza para el Desarrollo y la Cooperación COSUDE, CONDESAN, Helvetas Swiss Intercooperation, CDE University of Bern.
- Mazón M (2016) Taking shortcuts to measure species diversity: parasitoid Hymenoptera subfamilies as surrogates of species richness. Biodivers Conserv 25: 67–76. https://doi.org/10.1007/s10531-015-1029-y
- Mazón M, Aguirre N, Echeverría C, Aronson J (2019) Monitoring attributes for ecological restoration in Latin America and the Caribbean region. Rest Ecol 27(5): 992–999. https://doi.org/10.1111/rec.12986
- Mazón M, Bordera S (2008) Effectiveness of two sampling methods used for collecting Ichneumonidae (Hymenoptera) in Cabañeros National Park (Spain). European Journal of Entomology 105: 879–888. https://doi.org/10.14411/eje.2008.116
- Mazón M, Bordera S (2014) Diversity of Ichneumonidae (Insecta: Hymenoptera) in a protected area of Central Spain: what are we protecting? Insect Conserv Diver 7(5): 432–452. https://doi.org/10.1111/icad.12067

- Mazón M, Núñez-Penichet C, Cobos ME (2020) Relationship between body mass and forewing length in Neotropical Ichneumonidae (Insecta: Hymenoptera). Neotropical Entomology 49: 713–721. https://doi.org/10.1007/s13744-020-00784-9
- McGravy KW, Geroff RK, Gibbs J (2016) Malaise trap sampling efficiency for bees (Hymenoptera: Apoidea) in a restored tallgrass prairie. Florida Entomologist 99(2): 321–323. https://doi.org/10.1653/024.099.0230
- Michener CD (2007) The bees of the world. The Johns Hopkins University Press, Maryland.
- Montoya-Pfeiffer PM, Rodrigues RR, Alves dos Santos I (2020) Bee pollinator functional responses and functional effects in restored tropical forests. Ecological Applications 30(3): e02054. https://doi.org/10.1002/eap.2054
- Murcia C, Guariguata MR, Peralvo M, Gálmez V (2017) La restauración de bosques andinos tropicales: Avances, desafíos y perspectivas del futuro. Documentos Ocasionales 170, CI-FOR, Bogor.
- Myers N, Mittermeier RA, Mittermeier CG, da Fonseca GAB, Kent J (2000) Biodiversity hotspots for conservation priorities. Nature 403: 853–858. https://doi.org/10.1038/35002501
- Pandit R, Parrota J, Anker Y, Coudel E, Diaz Morejón CF, Harris J, Karlen DL, Kertész A, Mariño de Posada JL, Ntshotsho Simelane P, Tamin NM, Viera DLM (2018) Responses to halt degradation and to restore degraded land. In: Montanarella L, Scholes R, Brainich A (eds) The IPBES assessment report on land degradation and restoration. Secretariat of the Intergovernmental Science-Policy Platform on Biodiversity and Ecosystem Services, Bonn, Germany, 435–528.
- Pardo AS, Gonzalez PH (2007) Diversidad de abejas (Hymenoptera: Apoidea) en estados sucesionales del bosque húmedo tropical. Acta Biol Colomb 12(1): 43–56.
- Perillo LN, Neves FdS, Antonini Y, Martins RP (2017) Compositional changes in bee and wasp communities along Neotropical mountain altitudinal gradient. PLoS ONE 12(7): e0182054. https://doi.org/10.1371/journal.pone.0182054
- Polatto LP, Alves VV (2020) Bee species (Hymenoptera: Anthophila) in a Cerrado-Atlantic forest ecotone: nesting habits related to foraging activity in a degraded forest fragment. EntomoBrasilis 13: 0900. https://doi.org/10.12741/ebrasilis.v13.e0900
- Potts SG, Biesmeijer JC, Kremen C, Neumann P, Schweiger O, Kunin WE (2010) Global pollinator declines: trends, impacts and drivers. Trends Ecol Evol 25: 345–353. https://doi.org/10.1016/j.tree.2010.01.007
- Powney GD, Carvell C, Edwards M, Morris RKA, Roy HE, Woodcok BA, Isaac NJB (2019) Widespread losses of pollinating insects in Britain. Nature Research 10: 1018. https://doi.org/10.1038/s41467-019-08974-9
- Quintero E, Benavides AM, Moreno N, González-Caro S (2017) Bosques Andinos, estado actual y retos para su conservación en Antioquia. Medellín, Colombia: Fundación Jardín Botánico de Medellín Joaquín Antonio Uribe-Programa Bosques Andinos (COSUDE), Bogotá. Rasmussen C (2004) Abejas en el sur del Ecuador. Lyonia 7(2): 29–35.
- Requier F (2019) Bee colony health indicators: synthesis and future directions. CAB Reviews 14: 056. https://doi.org/10.1079/PAVSNNR201914056
- Reyes-Novelo E, Meléndez V, Delfín H, Ayala R (2009) Abejas silvestres (Hymenoptera: Apoidea) como bioindicadores en el Neotrópico. Trop Subtrop Agroecosystems 10(1): 1–13.

- Rossetti MR, Salvo A, Videla M, Valladares G (2013) Forest remnants contribute to parasitoid conservation: experimental evaluation of parasitism on a leafminer host. Journal of Insect Conservation 17: 1137–1144. https://doi.org/10.1007/s10841-013-9594-4
- Santos AMC, Quicke DLJ (2011) Large-scale diversity patterns of parasitoid insects. Entomol Sci 14: 371–382. https://doi.org/10.1111/j.1479-8298.2011.00481.x
- Santos Júnior JE, Ferrari RR, Nemésio A (2014) The orchid-bee fauna (Hymenoptera: Apidae) of a forest remnant in the southern portion of the Brazilian Amazon. Brazilian Journal of Biology 74(3): S184-S190. https://doi.org/10.1590/1519-6984.25712
- Saunders TE, Ward DF (2018) Variation in the diversity and richness of parasitoid wasps based on sampling effort. PeerJ 6:e4642. https://doi.org/10.7717/peerj.4642
- SER [Society for Ecological Restoration] (2004) The SER International primer on ecological restoration. Society for Ecological Restoration International, Tucson, Arizona.
- Solow AR (1993) A simple test for change in community structure. Journal of Animal Ecology 62: 191–193. https://doi.org/10.2307/5493
- Szejner P, Quesne CL, González ME, Christie DA (2011) Recuperación de bosques de *Pilgerodendron uviferum* (D. Don) Florin luego de perturbaciones antrópicas en la Cordillera de la Costa de Valdivia, Chile. Gayana Botanica 68: 196–206. https://doi.org/10.4067/S0717-66432011000200010
- Taki H, Murao R, Mitai K, Yamaura Y (2018) The species richness/abundance-area relationship of bees in an early successional tree plantation. Basic and Applied Ecology 26: 64–70. https://doi.org/10.1016/j.baae.2017.09.002
- Theodorou P, Radzeviciute R, Lentendu G, Kahnt B, Husemann M, Bleidorn C, Settele J, Schweiger O, Grosse I, Wubet T, Murray TE, Paxton RJ (2020) Urban areas as hotspots for bees and pollination but not a panacea for all insects. Nature Communications 11: 576. https://doi.org/10.1038/s41467-020-14496-6
- Townes H (1972) A light-weight Malaise trap. Entomol News 83: 239–247.
- Van Noort S (2004) Ichneumonid (Hymenoptera: Ichneumonoidea) diversity across an elevational gradient on monts Doudou in southwestern Gabon. California Academy of Sciences Memoir 28: 187–216.
- Veijalainen A, Sääksjärvi IE, Tuomisto H, Broad GR, Bordera S, Jussila R (2014) Altitudinal trends in species richness and diversity of Mesoamerican parasitoid wasps (Hymenoptera: Ichneumonidae). Insect Conservation and Diversity 7(6): 496–507. https://doi.org/10.1111/icad.12073
- Vieira LC, Oliveira NG, Brewster CC, Gayubo ST (2012) Using higher taxa as surrogates of species level data in three Portuguese protected areas: a case study of Spheciformes (Hymenoptera). Biodiversity and Conservation 21: 3467–3486. https://doi.org/10.1007/s10531-012-0374-3
- Volpato A, Ahmed KSD, Williams CD, Day MF, O'Hanlon A, Ruas S, Rotchés-Ribalta R, Mulkeen C, Huallacháin DO, Gormally MJ (2020) Using Malaise traps to assess aculeate Hymenoptera associated with farmland linear habitats across a range of farming intensities. Insect Conservation and Diversity 13(3): 229–238. https://doi.org/10.1111/icad.12383

- Vos M, Berrocal SM, Karamaouna F, Hemerik L, Vet LEM (2001) Plant-mediated indirect effects and the persistence of parasitoid-herbivore communities. Ecology Letters 4: 38–45. https://doi.org/10.1046/j.1461-0248.2001.00191.x..
- Widhiono I, Sudiana E, Darsono D (2017) Diversity of wild bees along elevational gradient in an agricultural area in Central Java, Indonesia. Psyche: A Journal of Entomology 2968414. https://doi.org/10.1155/2017/2968414
- Yepes AP, del Valle JI, Jaramillo SL, Orrego SA (2010) Recuperación estructural en bosques sucesionales andino de Porce (Antioquia, Colombia). Revista de Biología Tropical 58: 427–445. https://doi.org/10.15517/rbt.v58i1.5220